

Supplementary Figures and Tables

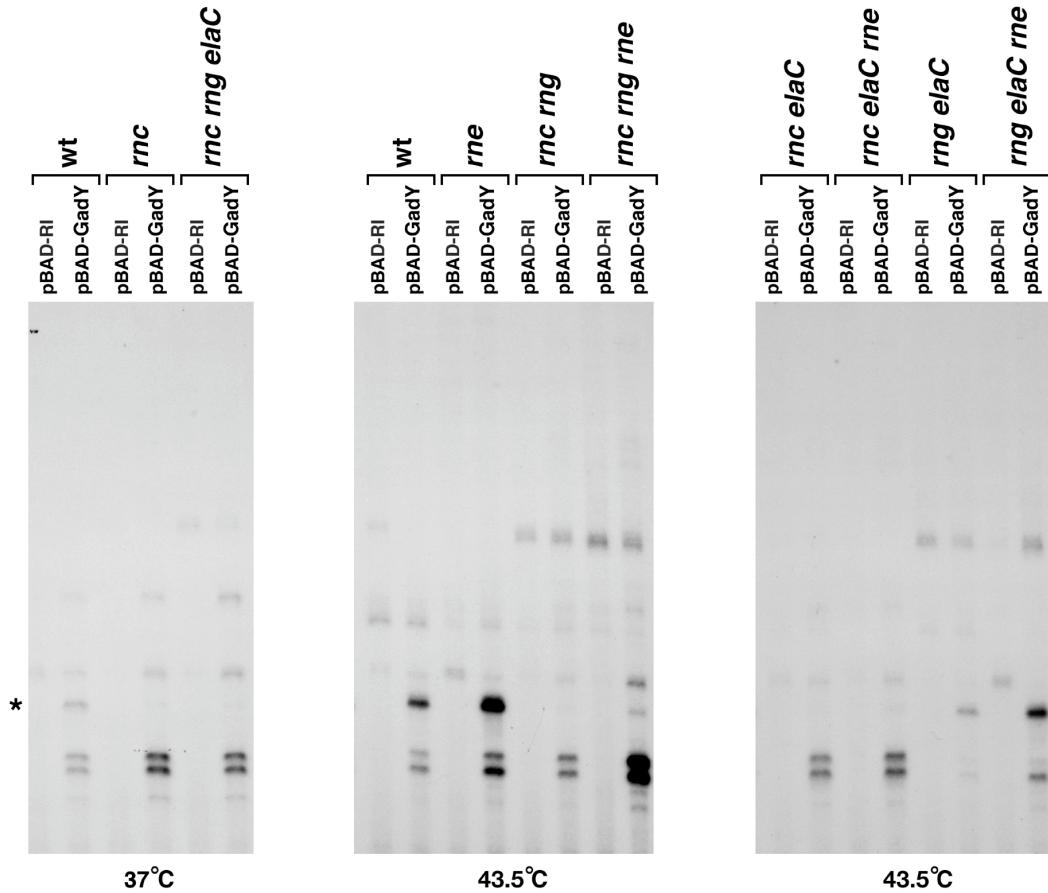
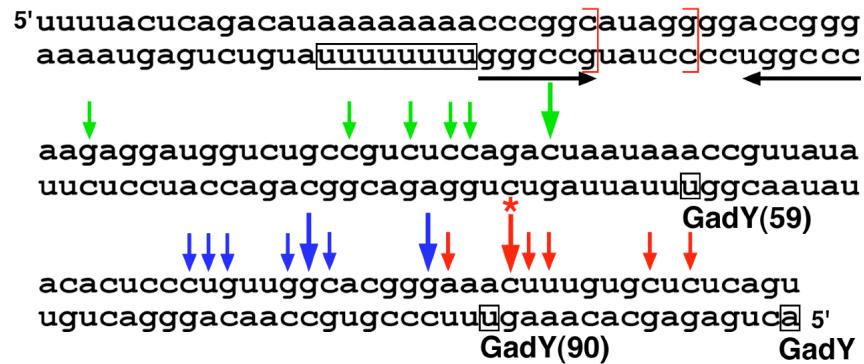


Fig. S1. GadY-directed processing in RNase triple mutant strains. Total RNA was isolated from wild type and RNase mutant derivatives of the *lacZ_{gadX}* reporter strain harboring pBAD-RI and pBAD-GadY, 20 min following the addition of arabinose to 0.2% to actively growing cultures ($OD_{600} \approx 0.7$). For the strains carrying the temperature sensitive *rne* alleles, the culture was shifted to the non-permissive temperature (43.5°C) for 30 min prior to induction. Samples were analyzed as in Fig. 4. The band decreased in *rnc* mutant strains is indicated by the asterisk, and the extra band present in *rng* mutant strains is indicated by the bullet.



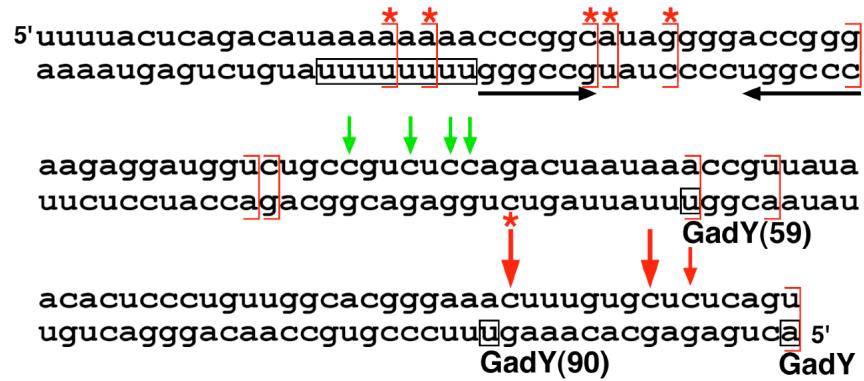


Fig. S3. 5'-ends detected by primer extension assays and 3'-ends detected by 3' RACE assays for products of in vivo cleavage in wild type and *rnc* mutant strains. 5' ends are indicated by arrow, 3' ends are indicated by brackets, and ends specific to the wild type strain are indicated by asterisks. Red denotes the GadY-dependent ends and green denotes the pRI-GadY(59)-dependent ends.

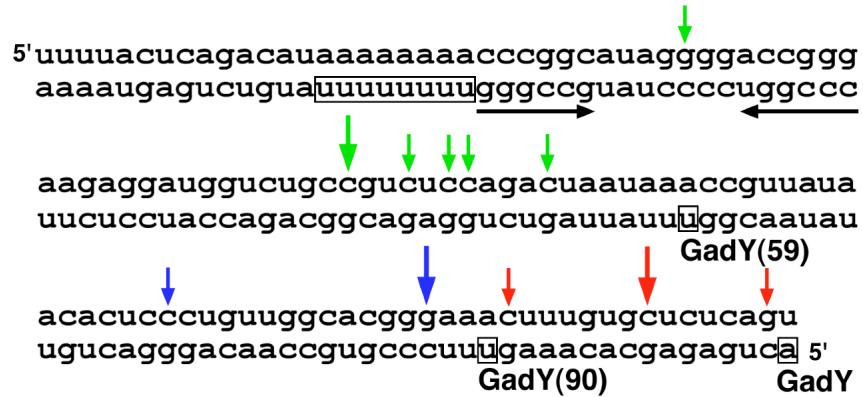


Fig. S4. 5'-ends detected by primer extension assays for products from in vitro assay with *rnc* mutant extracts. 5' ends are indicated by arrows. Red denotes the GadY-dependent ends, blue denotes the GadY(90)-dependent ends and green denotes the pRI-GadY(59)-dependent ends.

Table S1. Oligonucleotides used in study

Name	Sequence (5'-3', restriction sites underlined)	Use
GadY-S1	cacacaga <u>attt</u> ttagtctggagacggcagac	pRI-GadY(59), pBAD-GadY(59) PCR
GadY-A1	cacaca <u>aagctt</u> aaaaaaaaaccggcataggggacgg	pRI-GadY(59)
lacZ-S3	ccacaggat <u>ccggcc</u> gattcattaatgcagc	pACYC-lacZ PCR
lacZ-A3	cacaca <u>aagctttt</u> attttgacaccagacca	pACYC-lacZ PCR
gadW-A1	gcagcagaata <u>actctcg</u> ttatgcagcg	pACYC-lacZ _{gadW} PCR, gadW Northern detection
gadX-S1	ctagcccgaat <u>cgcc</u> cagega	pACYC-lacZ _{gadX} PCR
lacZ-S10	caca <u>aagatctt</u> ccaggta <u>cacgcgtt</u> gtaa	pACYC-lacZ _{gadX} PCR
lacZ-A6	cacact <u>cgaga</u> acctggcgta <u>cccaactt</u>	pACYC-lacZ _{gadX} PCR
gadY-QC1	ggaaggat <u>gggtctgcgc</u> tc	pACYC-lacZ _{gadX} PCR
gadY-QC2	gacggc <u>aggaccatcttcttcc</u>	pACYC-lacZ _{gadX} PCR
IG-S1	caca <u>aqatcttt</u> tact <u>cagacataaaaaa</u>	pACYC-lacZ _{gadX} PCR
IG-A2	cacact <u>cqagact</u> gagagcacaa <u>gttcc</u>	pACYC-lacZ _{gadX} PCR
AZ784	cgca <u>actctctactgttt</u> g <u>attt</u> c <u>accgc</u> tttttggcta	pBAD-RI PCR
AZ785	tagccaaaaaa <u>acgggt</u> g <u>attt</u> ca <u>aacagt</u> agaggttgcg	pBAD-RI PCR
JK116	gttat <u>attt</u> g <u>attt</u> act <u>gagac</u> ac	pBAD-GadY PCR
JK117	cttact <u>gagac</u> g <u>caattt</u> ttccgtgc	pBAD-GadY(90) PCR
JK118	gaata <u>aggattttaaqcttt</u> tact <u>catc</u> ag	pBAD-GadY, pBAD-GadY(90), pBAD-GadY(59) PCR
cat-S2	gtcat <u>cctgccc</u> ta <u>gaa</u> ac <u>atc</u> ac <u>at</u> at <u>catc</u> tc <u>gtt</u> tc <u>ccgc</u> tt <u>cg</u> ga <u>agat</u> ca <u>t</u> tc <u>cg</u> ca <u>gaaa</u>	GSO129 PCR
Px-cat-S1	ttaata <u>aaac</u> ag <u>taat</u> at <u>ttt</u> at <u>gt</u> ta <u>at</u> ta <u>at</u> ta <u>at</u> g <u>taact</u> at <u>gg</u> g <u>aaaaaa</u> at <u>cact</u> gg <u>atata</u> ac	GSO129 PCR
gfp-S3	gccggaa <u>atacc</u> cc <u>agg</u> c <u>ctc</u>	GSO403 PCR
gfp-A2	tgg <u>cagtcc</u> ct <u>act</u> ct <u>cg</u> agg <u>tc</u> ag <u>cta</u> at <u>aa</u> g <u>ctt</u>	GSO403 PCR
gfp-S2	taa <u>actcc</u> ct <u>gttgc</u> ac <u>ggaa</u> act <u>tttgc</u> tc <u>at</u> ca <u>gttgc</u> at <u>ccgtc</u> g <u>act</u> ct <u>tagagg</u>	GSO403 PCR
PS2-X-A1	at <u>gtttataaaaaaa</u> at <u>ggct</u> at <u>ttt</u> at <u>tttcc</u> ag <u>taaa</u> cat <u>at</u> ga <u>aat</u> at <u>cct</u> c <u>ttt</u> at <u>gt</u>	GSO403, GSO404 PCR
pKD4-S2	tgc <u>gagat</u> gg <u>ggaa</u> ct <u>gtcc</u>	GSO403, GSO404 PCR
pKD4-A2	gtt <u>gtgg</u> ac <u>aa</u> ac <u>agg</u> c <u>agg</u>	GSO403, GSO404 PCR
lacZ-234-R	tgg <u>cagtcc</u> ct <u>act</u> ct <u>cg</u> cat <u>tttgc</u> at <u>tttgc</u> at <u>tttgc</u>	GSO404 PCR
lacZ-S3	cacaggat <u>ccggcc</u> gatt <u>at</u> at <u>gc</u> ag <u>c</u>	GSO404 PCR
GadX-LacUP	tc <u>tc</u> tc <u>cc</u> gt <u>taa</u> ac <u>at</u> ac <u>ac</u> at <u>at</u> tc <u>c</u> tc <u>gtt</u> tc <u>cc</u> gt <u>tc</u> tc <u>cc</u> gt <u>ca</u> at <u>at</u> gt <u>g</u> at <u>tt</u> g <u>at</u> tc <u>c</u> ac	GSO404 PCR
PS1-rg	at <u>gacgg</u> ct <u>gta</u> tt <u>gtt</u> gt <u>taa</u> ac <u>gt</u> ta <u>ac</u> gc <u>c</u> tc <u>tc</u> tc <u>cc</u> gt <u>gt</u> at <u>gg</u> ct <u>gg</u> at <u>gt</u> tc <u>tc</u>	GSO410 PCR (Δrng)
PS2-rg	g <u>ccca</u> g <u>cat</u> ct <u>gtt</u> tt <u>at</u> at <u>cc</u> at <u>ac</u> tg <u>ac</u> gc <u>gt</u> caa <u>act</u> ct <u>gt</u> ct <u>ct</u> gc <u>at</u> at <u>gt</u> aa <u>at</u> tc <u>tc</u> tt <u>at</u> gt	GSO410 PCR (Δrng)
PS1-mlA	at <u>ttgt</u> ag <u>gtt</u> cc <u>ccat</u> at <u>gtt</u> ct <u>at</u> gg <u>at</u> cc <u>agg</u> a <u>gtt</u> gt <u>agg</u> ct <u>gg</u> ag <u>ct</u> gt <u>ctt</u> tc	GSO411 PCR ($\Delta rnlA$)
PS2-mlA	g <u>acac</u> gt <u>ttt</u> ag <u>g</u> cc <u>cc</u> tg <u>agac</u> gt <u>ta</u> at <u>cc</u> gg <u>gt</u> att <u>tc</u> at <u>at</u> ga <u>aa</u> at <u>c</u> tc <u>cc</u> tt <u>at</u> gt	GSO411 PCR ($\Delta rnlA$)
mh686	ta <u>tgc</u> tt <u>at</u> act <u>agcc</u> ga <u>at</u> tttt <u>gaa</u> ac <u>gt</u> at <u>g</u> a <u>act</u> ct <u>gt</u> tg <u>agg</u> ct <u>gg</u> ag <u>ct</u> gt <u>tc</u>	GSO412 PCR ($\Delta elacC$)
mh687	ata <u>acagg</u> gc <u>at</u> tt <u>cc</u> gt <u>tttt</u> ta <u>ataa</u> tg <u>acag</u> gt <u>ta</u> at <u>g</u> a <u>at</u> at <u>c</u> tc <u>cc</u> tt <u>at</u> gt	GSO412 PCR ($\Delta elacC$)
gadX-T7	ct <u>tg</u> at <u>tt</u> ta <u>at</u> ac <u>gact</u> ca <u>ct</u> at <u>at</u> gg <u>agag</u> at <u>tt</u> tc <u>ga</u> ac <u>cc</u> gt <u>tc</u> ac	gadX in vitro transcription template PCR
gadX-T7-R	gc <u>ac</u> ga <u>at</u> at <u>ttt</u> tc <u>gc</u> ac	gadX in vitro transcription template PCR
gadY-T7	ct <u>tg</u> at <u>tt</u> ta <u>at</u> ac <u>gact</u> ca <u>ct</u> at <u>at</u> gg <u>act</u> g <u>agag</u> ca <u>aa</u> at <u>ttt</u> cc	GadY in vitro transcription template PCR
gadY90-T7	ct <u>tg</u> at <u>tt</u> ta <u>at</u> ac <u>gact</u> ca <u>ct</u> at <u>at</u> gg <u>ttt</u> at <u>tt</u> at <u>gt</u> tc <u>gg</u> ga <u>ac</u> gg <u>ac</u> gc	GadY(90) in vitro transcription template PCR
gadY59-T7	ct <u>tg</u> at <u>tt</u> ta <u>at</u> ac <u>gact</u> ca <u>ct</u> at <u>at</u> gg <u>ttt</u> at <u>tt</u> at <u>gt</u> tc <u>gg</u> ga <u>ac</u> gc	GadY(59) in vitro transcription template PCR
gadY-T7-R	aaaa <u>aaaa</u> ac <u>cc</u> gc <u>at</u> agg <u>gg</u> ac <u>cc</u> gc	GadY, GadY(90), GadY(59) in vitro transcription template PCR
gfp-R	t <u>tttgg</u> cc <u>at</u> gg <u>aa</u> ac <u>agg</u> gt <u>tttcc</u> ag <u>t</u>	gfp Northern detection
gadX-A1	g <u>tttgc</u> at <u>cc</u> g <u>aaaa</u> ac <u>cc</u> ag <u>gt</u> tc <u>acc</u> gc <u>cc</u>	gadX Northern detection
cat-A1	agg <u>cc</u> gt <u>at</u> at <u>cc</u> ag <u>gt</u> tc <u>ga</u> ac <u>cc</u> gt <u>tc</u> gt	cat Northern detection
lacZ-R2	gg <u>cc</u> ct <u>tcc</u> gt <u>tta</u> tc <u>gc</u> ca <u>cc</u> gt <u>cc</u> ag <u>gt</u> gg <u>cg</u> ca	lacZ primer extension
GadW-A2	ac <u>ga</u> at <u>at</u> tt <u>tc</u> gc <u>ac</u> ag <u>gt</u> ta <u>at</u> tc <u>tt</u>	gadW primer extension
EF584	gg <u>tttatt</u> g <u>aga</u> at <u>at</u> gt <u>ttt</u> tc <u>gt</u> tc <u>ag</u> cc	cat 3' RACE
EF607	ct <u>ttgg</u> ac <u>aca</u> at <u>tt</u> gg <u>aa</u> ata <u>ca</u> act <u>ata</u> act <u>c</u>	gfp 3' RACE

Table S2. Plasmids used in study

Name	Resistance markers	Source or reference
pRI	(Amp ^R)	1
pRI-GadY	(Amp ^R)	1
pRI-GadY(90)	(Amp ^R)	This study
pRI-GadY(59)	(Amp ^R)	This study
pBAD18	(Amp ^R)	2
pBAD-RI	pBAD18 with an EcoRI site at the transcription start site (Amp ^R)	This study
pBAD-GadY	(Amp ^R)	This study
pBAD-GadY(90)	(Amp ^R)	This study
pBAD-GadY(59)	(Amp ^R)	This study
pACYC184	(Cm ^R , Tet ^R)	3
pACYC-lacZ	(Cm ^R)	This study
pACYC-lacZ _{gadX}	(Cm ^R)	This study
<i>placZ-gadX_{gadY-10}</i> mutant	(Cm ^R)	1
pKD3	(Amp ^R , Cm ^R)	4
pKD4	(Amp ^R , Kan ^R)	4
pCP20	(Amp ^R)	5
pMG47	(Amp ^R , Kan ^R)	6

Table S3. Strains used in study

Name	Relevant genotype	Source or reference
MG1655	F- lambda- ilvG- rfb-50 rph-1	lab stock
GSO109	MG1655 <i>gadY-10</i>	1
NM500	MG1655 mini-λ-tet	N. Majdalani
NM700	MG1655 $\Delta lacZnm3$, mini-λ-cm	N. Majdalani
AC22	<i>zce726::Tn10 rne3071</i>	7
NHY322	<i>zci-501::Tn10 (tet) rnpA49</i>	8
EK442	$\Delta gadXW::kan$	9
NB478	<i>rnc::cat</i>	10
HT115	$\Delta rnc::Tn10$ (tet)	11
MIC3009	<i>rnhA::cat</i>	12
GSO129	MG1655 $\Delta gadX::P_{car}cat, gadY-10$	This study
GSO403	MG1655 $\Delta gadX::P_{car}cat \Delta gadW::gfp-rrnB$ T1 T2	This study
GSO404	MG1655 $\Delta lacZ \Delta gadXY::P_{lac-lacZ_{gadX}}$	This study
GSO405	MG1655 $\Delta lacZ \Delta gadXY::P_{lac-lacZ_{gadX}}\Delta rnc::cat$	This study
GSO406	MG1655 $\Delta lacZ \Delta gadXY::P_{lac-lacZ_{gadX}} zce726::Tn10$	This study
GSO407	MG1655 $\Delta lacZ \Delta gadXY::P_{lac-lacZ_{gadX}} zce726::Tn10 rne3071$	This study
GSO408	MG1655 $\Delta lacZ \Delta gadXY::P_{lac-lacZ_{gadX}} zci-501::Tn10$	This study
GSO409	MG1655 $\Delta lacZ \Delta gadXY::P_{lac-lacZ_{gadX}} zci-501::Tn10 rnpA49$	This study
GSO410	MG1655 $\Delta lacZ \Delta gadXY::P_{lac-lacZ_{gadX}}\Delta rng::kan$	This study
GSO411	MG1655 $\Delta lacZ \Delta gadXY::P_{lac-lacZ_{gadX}}\Delta rnlA::kan$	This study
GSO412	MG1655 $\Delta lacZ \Delta gadXY::P_{lac-lacZ_{gadX}}\Delta elA::kan$	This study
GSO413	MG1655 $\Delta lacZ \Delta gadXY::P_{lac-lacZ_{gadX}}\Delta rnc::cat zce726::Tn10$	This study
GSO414	MG1655 $\Delta lacZ \Delta gadXY::P_{lac-lacZ_{gadX}}\Delta rnc::cat zce726::Tn10 rne3071$	This study
GSO415	MG1655 $\Delta lacZ \Delta gadXY::P_{lac-lacZ_{gadX}}\Delta rng::kan zce726::Tn10$	This study
GSO416	MG1655 $\Delta lacZ \Delta gadXY::P_{lac-lacZ_{gadX}}\Delta rng::kan zce726::Tn10 rne3071$	This study
GSO417	MG1655 $\Delta lacZ \Delta gadXY::P_{lac-lacZ_{gadX}}\Delta rnc::cat zci-501::Tn10$	This study
GSO418	MG1655 $\Delta lacZ \Delta gadXY::P_{lac-lacZ_{gadX}}\Delta rnc::cat zci-501::Tn10 rnpA49$	This study
GSO419	MG1655 $\Delta lacZ \Delta gadXY::P_{lac-lacZ_{gadX}}\Delta rnc::cat \Delta rng::kan$	This study
GSO420	MG1655 $\Delta lacZ \Delta gadXY::P_{lac-lacZ_{gadX}}\Delta rnc::cat \Delta rnlA::kan$	This study
GSO421	MG1655 $\Delta lacZ \Delta gadXY::P_{lac-lacZ_{gadX}}\Delta rnc::Tn10$	This study
GSO422	MG1655 $\Delta lacZ \Delta gadXY::P_{lac-lacZ_{gadX}}\Delta rnc::Tn10 \Delta rnhA::cat$	This study
GSO423	MG1655 $\Delta lacZ \Delta gadXY::P_{lac-lacZ_{gadX}}\Delta rnc::cat \Delta rng::kan zce726::Tn10$	This study
GSO424	MG1655 $\Delta lacZ \Delta gadXY::P_{lac-lacZ_{gadX}}\Delta rnc::cat \Delta rng::kan zce726::Tn10 rne3071$	This study
GSO425	MG1655 $\Delta lacZ \Delta gadXY::P_{lac-lacZ_{gadX}}\Delta rng::kan \Delta elA::cat zce726::Tn10$	This study
GSO426	MG1655 $\Delta lacZ \Delta gadXY::P_{lac-lacZ_{gadX}}\Delta rng::kan \Delta elA::cat zce726::Tn10 rne3071$	This study
GSO427	MG1655 $\Delta lacZ \Delta gadXY::P_{lac-lacZ_{gadX}}\Delta rnc::cat \Delta elA::kan zce726::Tn10$	This study
GSO428	MG1655 $\Delta lacZ \Delta gadXY::P_{lac-lacZ_{gadX}}\Delta rnc::cat \Delta elA::kan zce726::Tn10 rne3071$	This study
GSO429	MG1655 $\Delta lacZ \Delta gadXY::P_{lac-lacZ_{gadX}}\Delta elA::cat \Delta rnc::cat \Delta rng::kan$	This study
GSO430	MG1655 $\Delta gadXW::kan$	This study
GSO431	MG1655 $\Delta gadXW::kan \Delta rnc::cat$	This study
GSO432	MG1655 $\Delta gadX::P_{car}cat \Delta gadW::gfp-rrnB$ T1 T2 <i>rnc::Tn10-tet</i>	This study

References

1. Opdyke, J. A., Kang, J. G. & Storz, G. (2004). GadY, a small-RNA regulator of acid response genes in *Escherichia coli*. *J. Bacteriol.* **186**, 6698-6705.
2. Guzman, L. M., Belin, D., Carson, M. J. & Beckwith, J. (1995). Tight regulation, modulation, and high-level expression by vectors containing the arabinose P_{BAD} promoter. *J. Bacteriol.* **177**, 4121-4130.
3. Chang, A. C. Y. & Cohen, S. N. (1978). Construction and characterization of amplifiable multicopy DNA cloning vehicles derived from the P15A cryptic miniplasmid. *J. Bacteriol.* **134**, 1141-1156.
4. Datsenko, K. A. & Wanner, B. L. (2000). One-step inactivation of chromosomal genes in *Escherichia coli* K-12 using PCR products. *Proc. Natl. Acad. Sci. USA* **97**, 6640-6645.
5. Cherepanov, P. P. & Wackernagel, W. (1995). Gene disruption in *Escherichia coli*: Tc^R and Km^R cassettes with the option of Flp-catalyzed excision of the antibiotic-resistance determinant. *Gene* **158**, 9-14.
6. Batchelor, E. & Goulian, M. (2006). Imaging OmpR localization in *Escherichia coli*. *Mol. Microbiol.* **59**, 1767-1778.
7. Carpousis, A. J., Van Houwe, G., Ehretsmann, C. & Krisch, H. M. (1994). Copurification of *E. coli* RNAase E and PNPase: evidence for a specific association between two enzymes important in RNA processing and degradation. *Cell* **76**, 889-900.
8. Kirsebom, L. A., Baer, M. F. & Altman, S. (1988). Differential effects of mutations in the protein and RNA moieties of RNase P on the efficiency of suppression by various tRNA suppressors. *J. Mol. Biol.* **204**, 879-888.

9. Ma, Z., Richard, H., Tucker, D. L., Conway, T. & Foster, J. W. (2002). Collaborative regulation of *Escherichia coli* glutamate-dependent acid resistance by two AraC-like regulators, GadX and GadW (YhiW). *J. Bacteriol.* **184**, 7001-7012.
10. Yu, D., Ellis, H. M., Lee, E. C., Jenkins, N. A., Copeland, N. G. & Court, D. L. (2000). An efficient recombination system for chromosome engineering in *Escherichia coli*. *Proc. Natl. Acad. Sci. USA* **97**, 5978-5983.
11. Takiff, H. E., Chen, S. M. & Court, D. L. (1989). Genetic analysis of the *rnc* operon of *Escherichia coli*. *J. Bacteriol.* **171**, 2581-2590.
12. Itaya, M. & Crouch, R. J. (1991). A combination of RNase H (*rnh*) and *recBCD* or *sbcB* mutations in *Escherichia coli* K12 adversely affects growth. *Mol. Gen. Genet.* **227**, 424-432.